# CF33-CD19T ONCOLYTIC VIRUS (onCARIytics) IN COMBINATION WITH OFF-THE-SHELF ALLOGENEIC CYCART-19 T-CELLS TARGETING DE NOVO CD19T EXPRESSING TUMORS

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### Introduction

Autologous chimeric antigen receptor (CAR) T Cell therapy has shown impressive clinical responses against CD19+ B-Cell hematological malignancies and is being actively explored in the treatment of solid tumors. However, several barriers have precluded therapeutic responses in solid tumors, including limited tumor-restricted CAR targets and the immunosuppressive tumor microenvironment. We have recently reported the successful combination immunotherapy using a novel chimeric vacciniabased oncolytic virus (OV), called onCARlytics (Imugene Limited), that is engineered to express a non-signaling, truncated CD19 (CD19t) antigen for tumor-selective delivery, enabling de novo targeting of tumor cells by autologous CD19-CAR T Cell. One of the field's unanswered questions is whether treatment-naïve allogeneic CAR T Cell are superior to cancer patient-derived T-Cells for product manufacturing to improve overall responses against solid tumors.

Here, we evaluated this combination strategy using two allogeneic CAR T Cell products generated from peripheral blood mononuclear cells (PBMC) and placental T-Cells, respectively. PBMC-derived CAR T Cell were manufactured from normal, healthy donors. CYCART-19 (Celularity<sup>®</sup>, Inc.) Cells were derived from postpartum human placental T-Cells that are genetically modified to express the CD19-CAR followed by CRISPR-Cas9- mediated knockout of the endogenous TCR and expanded to produce multiple doses of allogeneic "off the shelf" treatment.

CYCART-19 T-Cells induced potent cytolytic activity against solid tumor cells infected with onCARlytics. Interestingly, while we observed comparable anti-tumor activity between PBMCderived CD19-CAR T Cells and CYCART-19, significant differences in cytokine secretion were detected. This warrants the possibility that the placental-derived CAR T product may elicit reduced CRS potential in patients with maintained or improved efficacy. This combination approach demonstrated impressive in vivo anti-tumor response in human tumor xenograft models. In summary, our results have demonstrated that further development of this combination immunotherapy for the potential treatment of a wide array of solid tumors is warranted.

### Figure 1

### Delivering truncated CD19t (CD19t) to tumor cells using oncolytic virus (OV) as a target for CD19-CAR T Cell.

onCARlytics selectively infect solid tumor cells and deliver truncated CD19 (CD19t) as a target for CD19-CAR T Cell.



### Figure 2

#### Postpartum human placental derived allogeneic **T-Cells expressing CAR-CYCART-19**

Celularity® has developed an allogeneic placental T-Cell with knockout of endogenous T-Cell receptors, derived from postpartum human placenta expressing CD19-CAR called CYCART-19. Placental-derived T-Cells are mostly naïve (CD45RA+ CCR7+), expand readily ex vivo, express markers of stem cell memory, and have lower expression of effector or exhaustion markers, which has been associated with greater stemness, enhanced proliferative capacity, and increased persistence in vivo.



### Figure 3

### Specific CYCART-19 tumor cell killing following onCARlytics infection

A Bright-field microscopy (10X magnification) of MDA-MB-468 tumor cells at 24h following onCARlytics infection or MDA-MB-468-CD19t (positive control lentivirally transduced to stably express CD19t) in the presence of untransduced (NT) or CYCART-19 T-Cells. B In vitro killing assay at 24h and C 48h of MDA-MB-468 tumor cells infected with onCARlytics and treated with untransduced autologous T-Cells, autologous CD19-CAR T Cell, NT (1 donor), or CYCART-19 (3 donors) T-Cells. Graph on the left represents tumor killing, and in the middle represents CD19t expression on tumor cells. Graph on the right represents tumor count against MDA-MB-468-CD19t treated with untransduced autologous T-Cells, autologous CD19-CAR T Cell, NT (1 donor), or CYCART-19 (3 donors) T-Cells.



### Figure 4

# infection

48h (right) by ELISA.





Figure 5





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## Activation of CYCART-19 by targeting of tumor cells expressing CD19t following onCARlytics

A Expression of activation marker (CD137) on untransduced autologous T-Cells, autologous CD19-CAR T Cell, untransduced [NT] (1 donor), or CYCART-19 (3 donors) T-Cells following 24h (left) and 48h (right) in vitro co-culture with MDA-MB-468 tumor cells infected with onCARlytics. 🖲 IFNγ and C IL-2 production following in vitro infection of MDA-MB-468 tumor cells with onCARlytics in the presence of autologous untransduced, autologous CD19-CAR, NT, or CYCART-19 T-Cell measured at 24h (left) and 48h (right) by ELISA. Ο IFNγ and IL-2 production following in vitro co-culture of MDA-MB-468-CD19t with autologous untransduced, autologous CD19-CAR, NT (1 donor), or CYCART-19 (1 donors) T-Cells measured at 24h (left) and

### CD19t expression in tumor cells following onCARlytics infection in vivo

Subcutaneously engrafted MDA-MB-468 tumors were collected 3, 7, or 10 days from NSG mice following onCARlytics infection at three indicated virus pfu per mouse and analyzed via flow cytometry for the expression of CD19t. MDA-MB-468 lentivirally transduced to stably express CD19t were used as a positive control (+ctrl).



### Schema of in vivo studies testing onCARlytics in combination with CYCART-19



### Comparing anti-tumor activity of CYCART-19 against autologous CD19-CAR T Cell in MDA-MB-468-CD19t bearing NSG mice

Mice were engrafted with subcutaneous MDA-MB-468-CD19t (5x10<sup>6</sup> cells) and were intravenously treated with untransduced autologous, autologous CD19-CAR, NT, CYCART-19 (2 donors) T-Cells (5x10<sup>6</sup> cells). Tumors were measured to determine T-Cell efficacy against a positive control tumor cell line in vivo.



### Figure 8

### Anti-tumor activity of CYCART-19 in combination with onCARlytics in human xenograft triple negative breast cancer tumor model

Mice were engrafted with subcutaneous MDA-MB-468 (5x10<sup>6</sup> cells) and were intratumorally treated with 0 or 10<sup>6</sup> pfu of onCARlytics per mouse. Mice were intravenously treated with untransduced autologous, autologous CD19-CAR, NT, CYCART-19 (2 donors) T-Cells (5x10<sup>6</sup> cells) A Lines represent tumor volumes of individual mice per group (n=5-10) and **B** average of each group.





CYCART-19 treatment 7 days post onCARlytics infection shows significant tumor regression compared to onCARlytics or T-Cells alone in a xenograft model of triple negative breast cancer.

### References

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